



Saccharomyces cerevisiae Host Cell Proteins

Immunoenzymetric Assay for the Measurement of *Saccharomyces cerevisiae* Host Cell Proteins Catalog # F135

Intended Use

This kit is intended for use in determining the presence of *Saccharomyces cerevisiae* protein contamination in products manufactured by recombinant expression in this yeast cell line. The kit is for Research and Manufacturing use only and is not intended for diagnostic use in humans or animals.

Summary and Explanation

Recombinant expression of proteins by *S. cerevisiae* is an efficient method to obtain cost effective quantities of a desired protein. Many of these recombinantly produced proteins are intended for use as therapeutic agents in humans and animals and as such must be highly purified. The manufacturing and purification process of these products leaves the potential for contamination by host cell proteins from *S. cerevisiae*. Such contamination can result in adverse toxic or immunological reactions and thus it is desirable to reduce host cell contamination to the lowest levels practical.

Immunological methods using antibodies to HCPs such as Western Blot and ELISA are widely accepted due to their specificity and sensitivity. While Western blot is a powerful method aiding in the identity of HCPs, it suffers from a number of limitations. Western blot is a complex and technique dependent procedure requiring a subjective interpretation of results. Furthermore, it is essentially a qualitative method and does not lend itself to obtaining quantitative answers. The sensitivity of Western blot is severely limited by the volume of sample that can be tested and by interference from the presence of high concentrations of the intended product. Western Blot may be able to detect HCPs in samples from upstream in the purification process but it often lacks adequate sensitivity to detect HCPs in purified downstream and final product. The microtiter plate immunoenzymetric assay (ELISA) method employed in this kit overcomes the limitations of Western blots providing on the order of 100 fold better sensitivity. This simple to use, highly sensitive, objective, and semi-quantitative ELISA is a powerful method to aid in optimal purification process development, process control, and in routine quality control and product release testing. This kit is "generic" in the sense that it

is intended to react with essentially all of the HCPs that could contaminate the product independent of the purification process. The antibodies have been generated against and affinity purified using a mild lysate washed of *S. cerevisiae* cells to obtain HCPs typically encountered in your initial product recovery step. Special procedures were utilized in the generation of these antibodies to insure that low molecular weight and less immunogenic contaminants as well as high molecular weight components would be represented. As such this kit can be used as a process development tool to monitor the optimal removal of host cell contaminants as well as in routine final product release testing. Because of the high sensitivity and broad reactivity of the antibodies, this generic kit has been successfully validated for testing of final product HCPs in many different products regardless of growth and purification process. When the kit can be satisfactorily validated for your samples, the application of a more process specific assay is probably not necessary in that such an assay would only provide information redundant to this generic assay. However, if your validation studies indicate the antibodies in this kit are not sufficiently reactive with your process specific HCPs it may be desirable to also develop a more process specific ELISA. This later generation assay may require the use of a more specific and defined antisera. Alternatively, if the polyclonal antibody used in this kit provides sufficient sensitivity and broad antigen reactivity, it may be possible to substitute the standards used in this kit for ones made from the contaminants that typically co-purify through your purification process and thus achieve better accuracy for process specific HCPs. The suitability of this kit for a given sample type and product must be determined and validated experimentally by each laboratory. The use of a process specific assay with more defined antigens and antibodies in theory may yield better sensitivity however such an assay runs the risk of being too specific in that it may fail to detect new or atypical contaminants that might result from some process irregularity or change. For this reason it is recommended that a broadly reactive "generic" host cell protein assay be used as part of the final product purity analysis even when a process specific assay is available.

Principle of the Procedure

The *Saccharomyces cerevisiae* Host Cell Protein assay is a two-site immunoenzymetric assay. Samples containing *S. cerevisiae* proteins are reacted in microtiter strips coated with an affinity purified anti-*S. cerevisiae* capture antibody. A second biotinylated anti-*S. cerevisiae* antibody is reacted simultaneously forming a sandwich complex of solid phase antibody-*S. cerevisiae* protein-biotin labeled antibody. The microtiter strips are then washed to remove any unbound reactants. Streptavidin: Alkaline Phosphatase is added to bind to any biotinylated antibody bound to the well. The substrate p-nitrophenyl phosphate (PNPP) is then reacted. The amount of hydrolyzed substrate is read on a microtiter plate reader and will be directly proportional to the concentration of *S. cerevisiae* proteins present.

Storage & Stability

- * All reagents should be stored a 2°C to 8°C for stability until the expiration date printed.
- * The substrate reagent should not be used if its absorbance at 405nm is greater than 0.4.
- * Reconstituted wash solution is stable until the expiration date of the kit

Reagents & Materials Provided

Component	Product #
Anti-<i>S. cerevisiae</i> Biotinylated Biotinylated affinity purified goat antibody in a protein matrix. 1x12mL	F136
Anti-<i>S. cerevisiae</i> coated strips 12x8 well strips in a bag with desiccant	F137
<i>S. cerevisiae</i> Standards Detergent solubilized proteins in a bovine serum albumin matrix. Standards at 0, 2, 8, 25, 75, & 200ng/mL. 1 mL/vial	F138
Streptavidin: Alkaline Phosphatase In a protein matrix with preservative, 1x12mL	F009
PNPP Substrate p-nitrophenyl phosphate in a Diethanolamine buffer, 1x12mL	F008
Wash Concentrate (20X) Tris buffered saline with preservative, 1x50mL	F004

Materials & Equipment Required But Not Provided

Microtiter plate reader spectrophotometer with dual wavelength capability at 405 & 492nm.
(If your plate reader does not provide dual wavelength analysis you may read at just the 405nm wavelength.)
Pipettors - 50µL and 100µL
Repeating or multichannel pipettor - 100µL
Microtiter plate rotator (150 - 200 rpm)
Sample Diluent (Recommend Cygnus Cat # I028)

Distilled water
1 liter wash bottle for diluted wash solution

Precautions

For research or manufacturing use only.
At the concentrations used in this kit none of the reagents are believed to be harmful. This kit should only be used by qualified technicians.

Preparation of Reagents

- * Bring all reagents to room temperature.
- * Dilute wash concentrate to 1 liter in distilled water, and label with kit lot and expiration date and store at 4°C.

Procedural Notes

1. Complete washing of the plates to remove excess unreacted reagents is essential to good assay reproducibility and sensitivity. We advise against the use of automated or other manual operated vacuum aspiration devices for washing plates as these may result in lower specific absorbances, higher non-specific absorbance, and more variable precision. The manual wash procedure described below generally provides lower backgrounds, higher specific absorbance, and better precision. If duplicate CVs are poor or if the absorbance of the 0 standard minus a substrate blank is greater than 0.300, evaluate plate washing procedure for proper performance.
2. High Dose Hook Effect may be observed in samples with very high concentrations of HCP. High Dose Hook Effect is due to insufficient excess of antibody for very high concentrations of HCPs present in the sample. Samples greater than 50 µg/mL may give absorbances less than the 200 ng/mL standard. It is also possible for samples to have certain HCPs in concentrations exceeding the amount of antibody for that particular HCP. In such cases the absorbance of the undiluted sample may be lower than the highest standard in the kit however these samples will fail to show acceptable dilutional recovery/parallelism as evidenced by an apparent increase in HCP concentration with increasing dilution. High Dose Hook is most likely to be encountered from samples early in the purification process. If a hook effect is possible, samples should also be assayed diluted. If the HCP concentration of the undiluted sample is less than the diluted sample this may be indicative of the hook effect. Such samples should be diluted at least to the minimum required dilution (MRD) where the dilution adjusted value remains essentially constant. The HCP value to be reported for such samples is the dilution corrected value at or greater than the established MRD. The diluent used should be compatible with accurate recovery. The preferred

diluent is our Cat# I028 available in 100mL, 500mL, or 1 liter bottles. This is the same material used to prepare the kit standards. As the sample is diluted in I028 its matrix begins to approach that of the standards thus reducing any inaccuracies caused by dilutional artifacts. Other prospective diluents must be tested for recovery by using them to dilute the 200ng/mL standard, as described in the “Limitations” section below.

3. If the substrate has a distinct yellow color prior to performing the assay it may have been contaminated. If this appears to be the case read 200 μ L of substrate against a water blank. If the absorbance is greater than 0.3 it may be necessary to obtain new substrate or the sensitivity of the assay may be compromised. The PNPP substrate is very sensitive to environmental contamination. Do not leave bottle open or at room temperature for longer than is needed. Only remove as much reagent as is needed for your assay run and do not return any unused substrate back into the substrate bottle. Additional substrate can be purchased separately as Cat # F008.

Limitations

* Before relying exclusively on this assay to detect host cell proteins, each laboratory should validate that the kit antibodies and assay procedure yield acceptable specificity, accuracy, and precision. A suggested protocol for this validation can be obtained by contacting our Technical Services Department or at our web site.

* The standards used in this assay are comprised of *S. cerevisiae* HCPs solubilized by mechanical disruption and detergent. Western blot analysis of the antibodies used in this kit demonstrates that they recognize the majority of distinct PAGE separated bands seen using a sensitive protein staining method like silver stain or colloidal gold. Because the vast majority of HCPs will be conserved among all strains of *S. cerevisiae* this kit should be adequately reactive to HCPs from your strain. Other clients have successfully validated this kit for their individual *S. cerevisiae* strains demonstrating acceptable specificity, accuracy, and sensitivity for process intermediate samples as well as final product. However, there can be no guarantee that this assay will detect all proteins or protein fragments from your process. In recognition of this potential limitation, we suggest reporting unknown samples in arbitrary dose units such as “ng/mL or parts per million of total immuno-reactive HCP equivalents”.

* Certain sample matrices may interfere in this assay. However the potential exists that the product protein itself, high or low pH, high salt, detergents, or other components in the sample matrix may result in either positive or negative interference in this assay. It is advised to test all sample matrices for interference by diluting the 200ng/mL standard, 1 part to 4 parts of the matrix containing no or very low HCP contaminants.

This diluted standard when assayed as an unknown should give a value of 30 to 50 ng/mL. Consult *Cygnus Technologies* Technical Service Department for advice on how to quantitate the assay in problematic matrices.

Assay Protocols

* The assay protocol is a simultaneous incubation of sample with biotinylated antibody. This yields a sensitivity of 0.5ng/mL and requires 4 hours to complete.

*The assay is very robust such that assay variables like incubation times, sample size, and other sequential incubation schemes can be altered to manipulate assay performance for more sensitivity, increased upper analytical range, or reduced sample matrix interference. Before modifying the protocol from what is recommended, users are advised to contact our technical services for input on the best way to achieve your desired goals.

*It is recommended that each laboratory assay appropriate quality control samples in each run to insure that all reagents and procedures are correct. You are strongly urged to make controls in your typical sample matrix using HCPs derived from your cell line. These controls can be aliquoted into single use vials and stored frozen for long-term stability.

* The protocol specifies the use of an approved microtiter plate shaker or rotator for the immunological step. If you do not have such a device it is possible to incubate the plate without shaking, however it will be necessary to extend the first immunological incubation step by one hour in order to achieve comparable results to the 2 hour shaking protocol. Do not shake during the 60 minute substrate incubation step as this may result in higher backgrounds and worse precision.

* Bring all reagents to room temperature.

* Set-up plate spectrophotometer to read dual wavelength at 405nm for the test wavelength and 492nm for the reference wavelength. Blank the instrument using the zero standard wells after assay completion.

* All standards, controls and test samples should be assayed at least in duplicate.

* Maintain a repetitive timing sequence from well to well for all assay steps to insure that all incubation times are the same for each well.

* Make a work list for each assay to identify the location of each standard control and patient.

Assay Protocol

1. Pipette 50µL of standards, controls and samples into wells indicated on work list.
2. Pipette 100µL of the biotinylated anti-*S. cerevisiae* (#F136) into each well.
3. Transfer to rotator and incubate at 180rpm for 2 hours.
4. Aspirate, or dump the contents to waste. Blot and vigorously bang out residual liquid over absorbent paper. Wash generously with diluted wash solution by flooding the wells with solution from a squirt bottle or by pipetting in ~350µL. Dump and bang again. Repeat for a total of 4 washes. Wipe off any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.
5. Pipette 100µL of Streptavidin: Alkaline Phosphatase (#F009) into each well.
6. Transfer to rotator at 180 rpm and incubate for 1 hour at room temperature, 24°C ± 4°.
7. Aspirate, or dump the contents to waste. Blot and vigorously bang out residual liquid over absorbent paper. Wash generously with diluted wash solution by flooding the wells with solution from a squirt bottle or by pipetting in ~350µL. Dump and bang again. Repeat for a total of 4 washes. Wipe off any liquid from the bottom outside of the microtiter wells as any residue can interfere in the reading step.
8. Pipette 100µL of PNPP substrate (#F008).
9. Incubate for 1 hour at room temperature.
10. Read absorbance at 405/492nm blanking on the Zero standard.

Calculation of Results

The standards may be used to construct a standard curve with values reported in ng/mL “total immunoreactive HCP equivalents” (See Limitations section above). This data reduction may be performed through computer methods using curve fitting routines such as

point to point, cubic spline, or 4 parameter logistic fit. **Do not use linear regression analysis to interpolate values for samples as this may lead to significant inaccuracies!** Data may also be manually reduced by plotting the absorbance values of the standard on the y-axis versus concentration on the x-axis and drawing a smooth point to point line. Absorbances of samples are then interpolated from this standard curve.

Example Data

Well #	Contents	Abs. at 405nm	Mean Abs.	ng/mL HCP equivs.
1A	Zero Std	0.000		
1B	Zero Std	0.004	0.002	
1C	2ng/mL	0.025		
1D	2ng/mL	0.023	0.024	
1E	8ng/mL	0.101		
1F	8ng/mL	0.095	0.096	
1G	25ng/mL	0.306		
1H	25ng/mL	0.295	0.300	
2A	75ng/mL	0.879		
2B	75ng/mL	0.865	0.872	
2C	200ng/mL	1.702		
2D	200ng/mL	1.722	1.712	
2E	sample A	0.099		
2F	sample A	0.096	0.098	8ng
2G	sample B	1.915		
2H	sample B	2.001	1.958	>200ng

Quality Control

- Precision on duplicate samples should yield average % coefficients of variation of less than 10% for samples greater than 2 ng/mL and < 200 ng/mL. CVs for samples < 2 ng/mL may be greater than 10%.
- For optimal performance the absorbance of the substrate when blanked against water should be < 0.4.
- It is recommended that each laboratory assay appropriate quality control samples in each run to insure that all reagents and procedures are correct.

Performance Characteristics

Cygnus Technologies has validated this assay by conventional criteria as indicated below. A more detailed copy of this “Validation Summary” report can be obtained by request. This validation is generic in nature and is intended to supplement but not replace certain user and product specific validation which should be performed by each laboratory.

Sensitivity

The lower limit of detection (LOD) is defined as that concentration corresponding to a signal two standard deviations above the mean of the zero standard. LOD is 0.5ng/mL.

The lower limit of quantitation (LOQ) is defined as the lowest concentration, where concentration coefficients of variation (CVs) are <20%. The LOQ is 0.9ng/mL.

Precision

Both intra (n=20 replicates) and inter-assay (n=5 assays) precision were determined on 3 pools with low (2ng/mL), medium (75ng/mL), and high concentrations (200ng/mL). The % CV is the standard deviation divided by the mean and multiplied by 100.

Pool	Intra assay CV	Inter assay CV
Low	15.4%	16.1%
Medium	4.2%	5.6%
High	2.1%	3.4%

Specificity/Cross-Reactivity

Immunoblot analysis against other strains of *S. cerevisiae* indicate that most of the proteins are conserved among all strains. Thus, this assay should be useful for detecting HCP's from other *Saccharomyces cerevisiae* strains. Cell proteins from the yeast *Pichia pastoris* were tested and found to be non-reactive.

Recovery/ Interference Studies

Various buffer matrices were evaluated by adding known amounts of *S. cerevisiae* HCP preparation used to make the standards in this kit. Because this assay is designed to minimize matrix interference most of these buffers yielded acceptable recovery defined as between 80-120%. In general extremes in pH (<5.0 and >8.5) as well as certain detergents can cause under-recovery. High concentrations of the recombinant product protein may also cause positive or negative interference in this assay. Each user should validate that their sample matrices yield accurate recovery. Such an experiment can be performed, by diluting the 200ng/mL standard provided with this kit, into the sample matrix in question and assaying for recovery.

Hook Capacity

Increasing concentrations of HCPs > 200 ng/mL were assayed as unknowns. The hook capacity, defined as that concentration which will give an absorbance reading less than the 200 ng/mL standard was >50 µg/mL.

Ordering Information/ Customer Service

To place an order or to obtain additional product information contact *Cygnus Technologies* Customer Support:

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